



The Role of p38 MAPK in Rheumatic Diseases

Athanasios Mavropoulos^{1,2} *PhD (UK)*, Andreas L. Koutsoumpas^{1,2,3} *MD*, Dimitrios P. Bogdanos^{1,2,3*} *MD*,

*PhD (UK)**, Lazaros I. Sakkas^{1,4} *MD,DM,PhD (UK)*

Abstract— p38 mitogen activated protein kinase (p38 MAPK) signaling appears to play a significant role in the regulation of immune-mediated inflammatory responses and therefore has been linked with several autoimmune diseases. This review discusses the current data regarding the involvement of p38 MAP in rheumatic diseases characterized by arthritis, with special attention in psoriatic arthritis, an arthritis with no apparent autoimmune features, and rheumatoid arthritis, an arthritis with apparent autoimmune features.

Keywords — autoimmunity, inflammation, psoriatic arthritis signaling, rheumatoid arthritis.

I. INTRODUCTION

Inflammation epitomizes an ordered sequence of events that establish and resolve a successful protective innate immune response against pathogens. Auto-inflammatory processes are sustained hyperactive immune response to self-antigens causing damage to tissues and organs [1-3]. Autoimmune rheumatic diseases (ARD) are heterogeneous, chronic auto-aggressive inflammatory disorders of the skin, connective tissue, and joints. ARD include among others rheumatoid arthritis, systemic sclerosis, and systemic lupus erythematosus [4]. Understanding the mechanisms responsible for the

regulation of inflammation is of paramount importance to better design targeted treatment strategies [5-8].

The innate immune system being quite sophisticated, can orchestrate the initiation, continuation, and cessation of inflammatory responses. Antigen presenting cells (APCs) engulf pathogens, present antigenic peptides to T cells and secrete monokines in order to regulate other innate immune cells such as neutrophils, dendritic cells, macrophages, monocytes and V γ 9/V δ 2 T cells [9, 10]. Yet, the role of certain innate participants, such as NK and NKT cells, is still largely undermined in ARD [11-14]. This occurs mainly due to their low-numbers in peripheral blood and tissues, which imposes a significant difficulty in their study. APC function has been recently attributed to NKs themselves and to certain hybrid subsets, such as natural killer dendritic cells (NKDCs), identified in mouse models and humans [15-17].

One of the most significant intracellular proteins that can initiate, perpetuate and resolve inflammation are mitogen-activated protein kinases (MAPKs) p38 primarily studied in macrophages [18, 19]. Accumulating evidence however suggest that p38 members are also activated within several innate cell subsets and may function as a critical players in ARD [20-23]. In this review we focus on two rheumatic diseases characterized by arthritis, psoriatic arthritis (PsA), which has no apparent autoimmune features, and rheumatoid arthritis (RA), which has apparent autoimmune features (rheumatoid factor, anti-citrullinated proteins antibodies [ACPAs]). ACPAs are considered pathogenic for RA [24].

P38 MAPK

Identification

p38 MAPK was originally described in 1994 as the mammalian homologue of the yeast Hog1 gene encoded kinase. In the same year it was also independently described as a kinase activated (phosphorylated) in response to an endotoxin (liposaccharide, LPS) and interleukin (IL-1) challenge, phosphorylating downstream targets, such as heat shock protein (hsp)27 and regulating inflammatory gene expression [25-27]. p38 is phosphorylated in response to inflammatory and stress stimuli, such as cytokines, ultraviolet irradiation, osmotic

Submitted Feb 23rd, 2015, accepted with revision Feb, 28, 2015.

This second paragraph will contain support information, including sponsor and financial support acknowledgment. For example, "Work performed by Andreas L. Koutsoumpas was has been co-financed by the European Union (European Social Fund-ESF) and Greek national funds through the Operational Program "Education and Lifelong Learning" of the National Strategic Reference Framework (NSRF)-Research Funding Program: "Heracleitus II, Investing in Knowledge society through the European Social Fund".

¹ Department of Rheumatology, Faculty of Medicine, School of Health Sciences, University of Thessaly, Biopolis, 41110 Larissa, Greece

² Cellular Immunotherapy and Molecular Immunodiagnosics, Institute of Research and Technology of Thessaly, Larissa 41222, Greece

³ Institute of Liver Studies, Transplantation Immunology and Mucosal Biology, King's College London School of Medicine at King's College Hospital, Denmark Hill Campus, London SE5 9RS, UK

⁴ Center for Molecular Medicine, Old Dominion University, Norfolk, VA, USA, 83024

*Correspondence to Dimitrios P. Bogdanos (e-mail: bogdanos@med.uth.gr).

shock, and heat shock, and is involved in cytokine regulation, cell differentiation and apoptosis [28].

Isoforms

Four subtypes of p38 proteins have been identified: α (MAPK14), β (MAPK11), γ (MAPK12/ERK6), and δ (MAPK13/SAPK4) [29-31]. p38 α and p38 β share approximately 75% gene sequence homology, whereas γ and δ are more distantly related, sharing just over 60% of their gene sequence. p38 α and p38 β are ubiquitously expressed whereas p38 γ and p38 δ show tissue-specific expression patterns. p38 α and p38 δ are abundantly expressed in macrophages, neutrophils and T cells, p38 γ is highly expressed in skeletal muscle and p38 β is abundant in endothelial cells [31, 32].

Phosphorylation

The structure of p38 kinases consists of a 135 amino acid N-terminal domain and a 225 amino acid C-terminal domain with the catalytic site located in the region linking the two domains [33]. p38 isoforms are shaped into different three-dimensional structures according to the precise orientation of the N- and C-terminal domains giving rise to variably sized ATP-binding pockets [34]. p38 members are phosphorylated within the ATP binding cleft on a single threonine (Thr-180) and a single tyrosine residue (Tyr-182). p38 kinases are phosphorylated (activated) after treatment with physiological (tumor necrosis factor [TNF]- α , IL-12 and IL-18, Toll-like receptor [TLR]-9, and TLR-4 ligands) and chemical stimuli, such as phorbol 12-myristate 13-acetate (PMA) plus ionomycin, sodium arsenite and anisomycin. Activation in different cellular compartments is dependent on the specificity of each stimulus.

Regulation

Since p38 phosphorylation can be induced by several agonists, the receptors and downstream converging signaling pathways diversify. Hence, studies have confirmed the existence of a classical activation pattern and two alternative ones [35, 36]. The classical MAPK pathway is evolutionarily conserved and encompasses a sequential phosphorylation of MAPK kinase kinases (MKKK) which activate a dual-specificity MAPK kinases (MKK), which in turn induce p38 MAPK by phosphorylating both threonine and tyrosine residues in a Thr-Xxx-Tyr motif [37, 38]. In the classical p38 MAPK cascade MEKK4 serves as an upstream MKKK and activates MKK3, MKK4, and then p38. MTK1, mixed lineage kinase (MLK) 2/3, apoptosis signal-regulating kinase (ASK) 1, and transforming growth factor

β -activated kinase (TAK) 1 are other MKKK kinases capable of activating MKK-6 or p38 signaling [39]. Further upstream activators of MKKKs include the growth arrest and DNA damage-inducible genes 45 (GADD45) proteins, that have been studied in detail and described to be of fundamental importance in leukocytes [40].

An increasing number of important substrates occur also downstream of p38. MAP kinase-activated protein kinase 2 (MK2) and MK3 are phosphorylated and can activate a variety of mediators, such as small HSP27, cAMP-response element-binding protein (CREB), and activating transcription factor (ATF) 1 [41, 42]. So far, various proteins have been identified as downstream substrates of p38, such as mitogen- and stress-activated kinase (MSK), p38-regulated/activated kinase (PRAK), and MAP kinase interaction protein kinase (MNK1) [43, 44]. Several novel proteins have also been identified in whole lysates of myoblasts as direct targets of p38 α , including Ahnak, Iws1, Grp78, Pgrmc, Prdx6, and Ranbp2 [45]. Moreover, TPL2/ERK1/2 kinases can also be regulated by p38 γ and δ isoforms [46].

Post-transcriptional regulation of gene expression by p38 MAPK is very important in the regulation of inflammatory responses [47]. Genome-wide analyses have demonstrated that only half of the alterations in gene expression during the immune response can be accounted for by transcriptional regulation. The rest are dependent on changes in mRNA stability [22, 48]. The modulation of mRNA stability is a powerful mechanism for bringing about rapid changes of gene expression, such as those that occur when the innate immune system first encounters a pathogen. Determinants of mRNA stability are usually located within the 3' untranslated region (UTR), and are tandem repeats of the sequence AUUUA termed adenosine/uridine-rich elements (AREs) [49, 50]. AREs contain, and are recognized by mRNA destabilizing proteins including tristetraprolin (TTP) and several others [51].

Phosphatases also exist to deactivate p38 so that p38 is not *ad infinitum* activated. Mitogen-activated protein kinase phosphatases (MKPs) can dephosphorylate MAPKs by binding to the TXY amino acid motif [52]. MKP-1, MKP-4, MKP-5, and MKP7 have been identified to effectively dephosphorylate p38 α and p38 β [53, 54]. MKPs however, are unable to dephosphorylate p38 γ or p38 δ [55].

P38 knock-outs

p38 α deficiency results in embryonic death due to defects in placental development and erythropoietin expression [56]. However, p38 β ^{-/-} mice are viable and exhibit no obvious defects in neither gene expression nor lymphocyte development [57]. Single knockouts of either



p38 γ or p38 δ , and the double knockout are also viable. Importantly, diminished expression of TNF- α , IL-1 β , and IL-10 was reported in stimulated macrophages isolated from p38 γ/δ null mice, which suggests that p38 γ/δ can be crucial regulatory constituents of the innate immune response [46].

Inhibitors

Dissection of the p38 MAPK signaling cascade is made possible with the development of specific p38 inhibitors [58, 59]. An array of pyridinyl imidazole anti-inflammatory compounds, such as SKF-86002, SB203580, and SB202190 were among the first available p38 inhibitors acting through competition for the ATP-binding site [60-63]. Pharmaceutical industries have invested heavily in developing competitive and specific compounds and a number of second and third generation of inhibitors, such as SC-79659, SC-80036, VXs, AMG-548, ML3403, pamapimod and AS1940477 [64, 65]. Most of these inhibitors show specific and strong activity against p38 and inhibit the production of proinflammatory cytokines, and, therefore, hold promise as therapeutic agents for chronic inflammatory diseases. For example pamapimod reduced clinical signs of arthritis, bone loss and inhibited TNF- α production in RA synovial explants [66]. Other novel compounds, such as GSK-681323, have been used to treat RA, SCIO-469 to treat multiple myeloma and dental pain, and RWJ67657 has been used as a broad anti-inflammatory agent [67]. Currently, efficacy and safety of p38 inhibitors are under evaluation in clinical trials [68].

p38 MAPK in psoriasis and psoriatic arthritis

The p38 MAPK pathway has been implicated in the pathogenesis of psoriasis, as it is detected by immunohistochemistry and Western blotting in psoriatic skin lesions [69, 70]. Kinase assays also confirmed the increased activity of p38 α , p38 β and p38 δ isoforms in lesional compared to non-lesional psoriatic skin. Phosphorylated p38 in lesional psoriatic epidermis, exhibited a distinct nuclear localization indicative of the kinase participation in the induction of active gene expression [70]. Dual specificity phosphatase (DUSP)1 that can dephosphorylate p38 MAPK and cease its function, is also impaired in psoriasis since its mRNA expression was significantly down-regulated in lesional compared to non-lesional psoriatic skin [71, 72].

Further downstream targets of p38 MAPK signaling, such as MK2, are also activated in the psoriatic epidermis [73]. Keratinocytes transfected with MK2-specific small interfering RNAs showed diminished MK2 expression and significant reduction in the expression of IFN- γ , TNF- α ,

IL-6, and IL-8 proteins. The mechanism by which p38 MAPK mediates its regulatory effects through downstream kinases has been dissected in mice with deleted MK2 [74]. These mice are deficient in the LPS-induced biosynthesis of several pro-inflammatory cytokines regulated by p38, including TNF- α , IFN- γ , IL-6, and IL-1. They survive LPS-induced endotoxic shock due to a reduction of the secretion of TNF- α by almost 90% [75]. MK2 has been considered as a key molecule participating in host defense against intracellular bacteria through regulation of both TNF- α and IFN- γ production [76, 77].

Mitogen- and stress-activated protein kinase 1 (MSK1) is another downstream target of p38 which regulates the expression of pro-inflammatory cytokine genes through activation of transcription factors. Western blotting analysis revealed a consistent and significant increase in phosphorylated MSK1 (Ser376) in lesional psoriatic skin [78, 79]. Cultured human keratinocytes incubated with anisomycin or IL-1 β resulted in the phosphorylation of both p38 MAPK and MSK1 (Ser376) whereas MSK1 (Ser376) phosphorylation was inhibited by pre-incubation with p38 inhibitors or dimethylfumarate [80]. In addition, transcription factors, such as cAMP/calcium responsive element binding protein (CREB) associated with cellular proliferation gene expression, are also phosphorylated in psoriatic skin [81]. Activation of CREB through ERK1/2 is directly linked with the expression of TNF- α , IL-6 and IL-8 [82]. These cytokines are also under direct regulation of the p38 pathway as well [83, 84]. p38 MAPK-induced phosphorylation of STAT-3 and of STAT-1 at serine 727 has also been demonstrated in lesional psoriatic skin [85, 86]. STAT-3, in particular, has been described as the crucial link between activated keratinocytes and immunocytes required for the development of psoriasis in a novel transgenic mouse model [87].

Thus, keratinocytes in the psoriatic epidermis are characterized not only by abnormal proliferation and apoptosis, but also by increased expression of inflammatory cytokines through interaction with immunocytes [84]. This seems to be regulated by the same signals arising from the activation of MAPK signaling cascades of p38 in immune cells [88]. Apart from the classical Th1 mediated response, Th17 cells have recently been demonstrated to sustain inflammation in psoriasis and psoriatic arthritis [89, 90]. The antimalarial drug and autophagy/lysosome inhibitor chloroquine (CHQ) is suggested as potential trigger of drug-induced psoriasis, in which Th17 cell mediated cytokine expression occurs in a p38-dependend IL-23 expression manner [91]. Dendritic cells treated with LPS increase the secretion of both IL-1 β and IL-23, and stimulate the secretion of IL-17, IFN- γ , and IL-22 by innate $\gamma\delta$ T cells [92]. IL-12, IL-23 and IL-27

production form a activation loop involving cells of innate and adaptive immune response [93]. In bone marrow-derived dendritic cells IL-23 and IL-27 expression is in part regulated by p38 MAPK [94].

Apart from inflammatory gene expression, antimicrobial peptide S100A8, known to be up-regulated in lesional psoriatic skin, was found also to be regulated by a p38 MAPK-dependent mechanism [95]. Similarly, p38-dependent expression was demonstrated for the antimicrobial peptides cathelicidin, human β -defensin-2, human β -defensin-3, and S100A7 in human keratinocytes [96].

Information on the involvement of MAPK signaling in the pathogenesis of PsA is very scarce, whereas activation of MAPKs, specifically p38 and downstream MK2, has been described in RA synovium and in the collagen-induced arthritis model of RA [64, 75]. Recently, in a mouse model it has been shown that psoriatic skin inflammation facilitated the development of arthritis and enthesitis, both features of PsA [97]. Th1 and Th17 Inflammatory cytokines are up-regulated in psoriasis, PsA and other spondyloarthritides [98-100] and TNF- α inhibitors are the mainstay treatment for psoriasis and PsA [101-103]. Back in 2000, Danning et al have linked elevated pro-inflammatory cytokines with NF κ B activation in PsA synovium [104]. More recent findings underlined the participation of both MAPK signaling and NF κ B activation in PsA synovium before and after treatment with TNF α inhibitor (etanercept) [105]. Activated p38 was present in both lining and sub-lining area of the synovial membrane and p38 positive cells were detected in inflammatory infiltrates and perivascular areas. In addition, IL-36 α is up-regulated in PsA and RA synovium and leads to IL-6 and IL-8 production by synovial fibroblasts through p38/NF κ B activation [106].

p38 MAPK pathway in rheumatoid arthritis

The p38 MAPK signaling pathway has been implicated in the pathogenesis of RA. In fact, for several years it has been regarded as a potential therapeutic target for RA and other chronic immune-mediated inflammatory diseases [28, 64].

RA is considered an autoimmune disease. Recent findings suggest that ACPAs are autoantigens in RA recognized by T cells and B cells and most importantly are arthritogenic [24]. Early studies have confirmed the heavy infiltration with T cells carrying activation markers in inflamed RA synovial membrane [107-109]. Infiltrating T

cells preferentially expressed IFN- γ , thus aggravating chronic inflammation. Local synthesis of IFN- γ in RA joints is largely induced by the synergistic effect of IL-12 and IL-18 that are produced by activated antigen presenting cells (APCs) [110, 111]. IL-12 and IL-18 induce IFN- γ through the p38 MAPK pathway in T and NK cells [22, 112]. Autoreactive T cells also maintain a spontaneous TNF- α production in rheumatoid synovial tissues via cell contact and cytokine activation [113]. Activation of p38 MAPK is a critical step for the acquisition of effector function in T cells [114, 115]. TNF- α is one of the major pro-inflammatory mediators in RA and recently, prolactin receptor (PRLR) which regulates the expression and release of TNF- α from CD14(+) monocytes through p38 MAPK was found to be markedly increased in RA patients [116]. This TNF- α release from CD14(+) monocytes can be abolished by PRLR gene silencing or treating with MAPK inhibitor.

Over the last few years, the importance of IL-17 and Th17 cells in the pathogenesis of RA has become apparent [117, 118]. Recent data suggest that engagement of TLR2 enhances IL-17(+) autoreactive T cell responses via p38 MAPK signaling in dendritic cells [119]. Inhibition of p38 MAPK activity dramatically decreased IL-17 gene expression and antigen-specific Th17 responses. In addition, increased salt solutions, found locally under physiological conditions, have been shown to activate the p38/MAPK pathway involving nuclear factor of activated T cells 5 (NFAT5); and serum/glucocorticoid-regulated kinase 1 (SGK1) during cytokine-induced Th17 polarization. In addition, increased dietary salt aggravated experimental autoimmune encephalomyelitis through Th17 cells [120].

Of the p38 MAPK isoforms p38 α is the one that strongly expressed in RA synovial tissue and mostly implicated in the pathogenesis of RA [66, 121]. Preferential activation of upstream MKK3 or MKK6 leading to p38MAPK activation in synovial fibroblasts has also been described [122, 123]. Deletion of p38 β does not affect experimental arthritis, whereas deletion of the Mapk14 (p38 α) confers protection against inflammatory bone loss. In the rat streptococcal cell wall arthritis model, inhibition of p38 MAPK leads to significant reduction of inflammation and cartilage breakdown [124]. Similar findings are also reported in other murine arthritic models, such as adjuvant-induced arthritis and collagen-induced arthritis [125, 126]. Animal models enable the prediction and testing of therapeutic targets, but the pathogenetic mechanisms involved may not be quite similar to those in human RA. Still, studies in animal models suggest that p38 MAPK and upstream kinase inhibition controls well-established arthritis [127]. In these models, several

early and new generation p38 MAPK inhibitors have been proven to be effective in reducing the disease severity.

Disappointingly enough, none of p38 inhibitors have successfully passed late clinical trials for the treatment of RA thus far [68]. Untoward effects that limited their use was significant liver toxicity, and their undesirable ability to cross blood-brain barrier [128]. Therefore, certain early inhibitors such as VX-745 (with promising results in phase II trials for RA) were soon discontinued [129]. Newer inhibitors, such as SCIO469, BIRB796, pamapimod, KR-00348 and AS1940477, are more selective at targeting p38, but show transient inhibition of pro-inflammatory markers [130, 131].

In order to explain the apparent failure of p38 inhibitors in RA, it is important to better understand the complexities of the p38 pathway and its communication with other cellular signaling pathways [28]. Targeting the upstream or MAP kinase kinases, may provide a viable alternative [127]. For example, MKK3 and MKK6, has been shown to partially maintain p38-mediated anti-inflammatory responses in bone marrow-derived macrophages (BMDM) [132]. Conventional p38 α inhibitors have limited efficacy in RA, possibly because p38 blockade suppresses the counter-regulatory mechanisms that limit inflammation. For instance, recent data revealed the existence of CD8+FoxP3+ Treg cells in peripheral blood of patients with RA [133]. Unlike their CD4+ counterparts, CD8+FoxP3+ Treg cells inhibited Th17 inflammatory responses thereby limit a wider range of inflammatory pathways. CD8+FoxP3+ Treg cell induction was supported both by p38 phosphorylation intrinsic to naive CD8+ T cells and by monocytes via CD86 and membrane TNF α . In contrast to conventional CD4+ T cells, freshly isolated natural Tregs exhibit marked activation of p38 MAPK [134]. The p38 MAPK pathway is also involved in the conversion of naive T cells into induced Tregs [135]. Inhibition of p38 MAPK activity prevents the TGF β -dependent conversion of CD4+CD25-T cells into Foxp3+ iTreg *in vitro* [136]. It follows that, p38 MAPK targeting (such as that through pharmacological inhibitors) potentially prevents the induction of iTreg *in vivo* and this may account for the inability of these antagonists *per se* to control the exacerbation of autoimmune disease.

All these data collectively define a largely unknown p38-dependent mechanism of FoxP3+ Treg cell induction. It seems that inhibition of p38 MAPK through pharmacological antagonists exerts two distinct functions. First, they act as potent anti-inflammatory agents by diminishing p38 MAPK mediated IFN- γ , IL-6, and IL-17 expression and; second they lead to functional impairment of suppressor cells, such as regulatory T and probably B

cells. While the former function of p38 MAPK antagonists is undisputed, the latter is largely unexplored but certainly unwanted.

Peripheral blood signature studies are of great importance and can provide a lot of information regarding new immune regulatory molecules [137]. Meticulous assessment of cellular populations their signaling pathways and associated gene expressions is necessary in order to advance our knowledge on the pathogenesis of rheumatic diseases and to successfully identify novel molecular therapeutic targets. Application of optimized protocols based on sensitive phospho flow cytometry have been recognized as promising alternatives for the investigation of the phosphorylation of p38 MAPK within different peripheral blood mononuclear cell (PBMC) populations [138-140]. For instance, we have optimized a flow cytometry-based assay that details cellular phenotypic status, signaling status and gene expression analysis, all combined [141, 142]. The availability of multiplexing technologies capable of simultaneously quantifying multiple biomarkers has been particularly helpful in dissecting changes in soluble cytokine and chemokine networks in clinical samples. (Fig. 1). Flow cytometry has been so far useful in revealing the phenotype of rare cells infiltrating psoriatic lesion and their secreted cytokines [143]. One such study in PBMC of patients with PsA identified a unique gene expression signature of MAPK signaling members [144]. These observations could be carefully projected and analyzed in relation to infiltrating lymphocytes from psoriatic skin biopsies [145, 146].

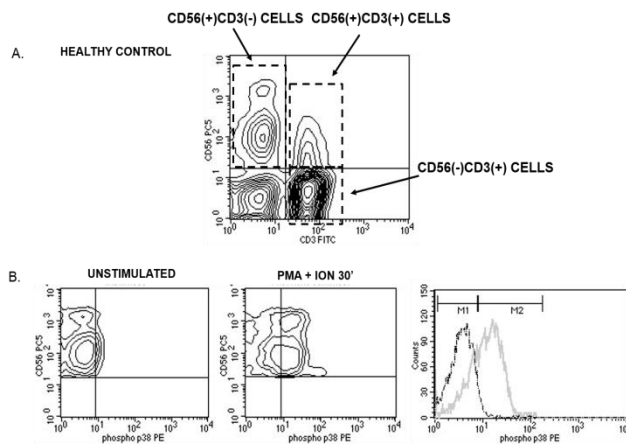


Fig.1 Flow cytometric analysis of P38 MAPK. Representative flow cytometric analysis of p38 MAPK phosphorylation in NK (CD56 positive) cells from a healthy donor following stimulation with PMA and Ionomycin

II. CONCLUSIONS

Current data suggest that p38 MAPK plays an important role in the pathogenesis of RA and possibly PsA. However, more studies are needed to further advance our knowledge on p38MAPK expression in inflammatory as well in suppressor cells during the course of arthritis in order to better define therapeutic strategies.

ACKNOWLEDGMENTS

We thank Dr Eirini I. Rigopoulou for critical reading of the manuscript.

Conflict of Interest: None.

REFERENCES AND FOOTNOTES

REFERENCES

- [1.] Perricone C, Toubi E, Valesini Gand Shoenfeld Y. Autoinflammation and autoimmunity: pathogenic, clinical, diagnostic and therapeutic aspects. *Isr Med Assoc J* **2014**;16(10):601-4.
- [2.] Cantarini L, Imazio M, Brizi MG, Lucherini OM, Brucato A, Cimaz Rand Galeazzi M. Role of autoimmunity and autoinflammation in the pathogenesis of idiopathic recurrent pericarditis. *Clin Rev Allergy Immunol* **2013**;44(1):6-13.
- [3.] Wahren-Herlenius Mand Dorner T. Immunopathogenic mechanisms of systemic autoimmune disease. *Lancet* **2013**;382(9894):819-31.
- [4.] Ceeraz S, Nowak EC, Burns CMand Noelle RJ. Immune checkpoint receptors in regulating immune reactivity in rheumatic disease. *Arthritis Res Ther* **2014**;16(5):469.
- [5.] Klareskog L, Gregersen PKand Huizinga TW. Prevention of autoimmune rheumatic disease: state of the art and future perspectives. *Ann Rheum Dis* **2010**;69(12):2062-6.
- [6.] Sakkas LI, Simopoulou T, Katsiari C, Bogdanos Dand Chikanza IC. Early systemic sclerosis-opportunities for treatment. *Clinical rheumatology* **2015**.
- [7.] Helmick CG, Felson DT, Lawrence RC, Gabriel S, Hirsch R, Kwok CK, Liang MH, Kremers HM, Mayes MD, Merkel PA, Pillemer SR, Reveille JD, Stone JHand National Arthritis Data W. Estimates of the prevalence of arthritis and other rheumatic conditions in the United States. Part I. *Arthritis Rheum* **2008**;58(1):15-25.
- [8.] Lawrence RC, Felson DT, Helmick CG, Arnold LM, Choi H, Deyo RA, Gabriel S, Hirsch R, Hochberg MC, Hunder GG, Jordan JM, Katz JN, Kremers HM, Wolfe Fand National Arthritis Data W. Estimates of the prevalence of arthritis and other rheumatic conditions in the United States. Part II. *Arthritis Rheum* **2008**;58(1):26-35.

Yet, there is very little information available on the kinases and signaling pathways activated in the rare NK and NKT cells of PsA patients (Fig. 2). As shown in figure we are currently able to successfully detect several surface epitopes together with phosphorylated kinases such as p38 and Stats and subsequent intracellular gene expression. We have previously shown that p38 MAPK regulates post-transcriptional IFN- γ gene expression in human NK and NKT cells [22]. This possibly occurs via an MKK6/p38/MK2-dependent mechanism for the stabilization of IFN- γ mRNA in NK and NKT cells, and may play an important role in host defense as well. We have previously applied the optimized methodology for the successful application of phospho-specific flow cytometry in order to detect phosphorylated p38 MAPK within peripheral and intrahepatic innate immune cells, such as NK and NKT [147]. Because of the critical and bi-directional role of the p38 MAPK / IFN- γ axis in inflammation and autoimmunity, the elucidation of the molecular mechanisms controlling its expression is the focus of ongoing research by our team [142].

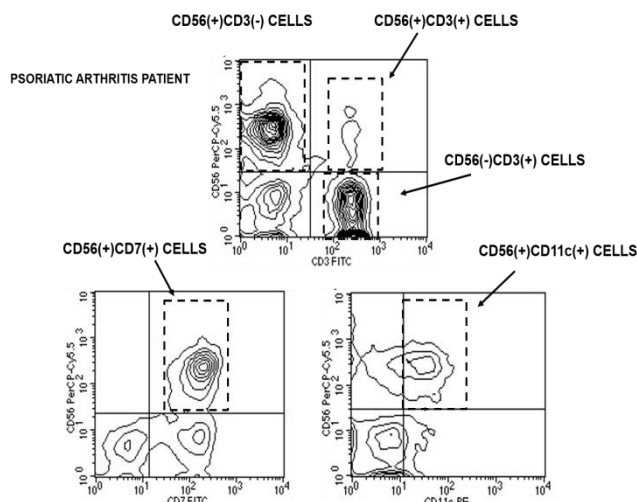


Fig 2. Representative histograms of experiments showing activation of p38 MAPK in a patients with psoriatic arthritis. Surface markers staining includes CD3 (pan-T marker), CD56 and CD7 (NK markers).



- [9.] Tyler CJ, Doherty DG, Moser Band Eberl M. Human Vgamma9/Vdelta2 T cells: Innate adaptors of the immune system. *Cell Immunol* **2015**.
- [10.] Mok MY. Tolerogenic dendritic cells: role and therapeutic implications in systemic lupus erythematosus. *Int J Rheum Dis* **2014**.
- [11.] Tobin AM, Lynch L, Kirby Band O'Farrelly C. Natural killer cells in psoriasis. *J Innate Immun* **2011**;3(4):403-10.
- [12.] Hervier B, Beziat V, Haroche J, Mathian A, Lebon P, Ghillani-Dalbin P, Musset L, Debre P, Amoura Zand Vieillard V. Phenotype and function of natural killer cells in systemic lupus erythematosus: excess interferon-gamma production in patients with active disease. *Arthritis Rheum* **2011**;63(6):1698-706.
- [13.] Issazadeh-Navikas S. NKT cell self-reactivity: evolutionary master key of immune homeostasis? *J Mol Cell Biol* **2012**;4(2):70-8.
- [14.] Shimizu K, Sato Y, Shinga J, Watanabe T, Endo T, Asakura M, Yamasaki S, Kawahara K, Kinjo Y, Kitamura H, Watarai H, Ishii Y, Tsuji M, Taniguchi M, Ohara Oand Fujii S. KLRG+ invariant natural killer T cells are long-lived effectors. *Proc Natl Acad Sci U S A* **2014**;111(34):12474-9.
- [15.] Lee SW, Park HJ, Kim Nand Hong S. Natural Killer Dendritic Cells Enhance Immune Responses Elicited by alpha -Galactosylceramide-Stimulated Natural Killer T Cells. *Biomed Res Int* **2013**;2013:460706.
- [16.] Pillarisetty VG, Katz SC, Bleier JI, Shah ABand Dematteo RP. Natural killer dendritic cells have both antigen presenting and lytic function and in response to CpG produce IFN-gamma via autocrine IL-12. *J Immunol* **2005**;174(5):2612-8.
- [17.] Plitas G, Chaudhry UI, Kingham TP, Raab JRand DeMatteo RP. NK dendritic cells are innate immune responders to *Listeria monocytogenes* infection. *J Immunol* **2007**;178(7):4411-6.
- [18.] Tudor C, Marchese FP, Hitti E, Aubareda A, Rawlinson L, Gaestel M, Blackshear PJ, Clark AR, Saklatvala Jand Dean JL. The p38 MAPK pathway inhibits tristetraprolin-directed decay of interleukin-10 and pro-inflammatory mediator mRNAs in murine macrophages. *FEBS Lett* **2009**;583(12):1933-8.
- [19.] Saklatvala J. The p38 MAP kinase pathway as a therapeutic target in inflammatory disease. *Curr Opin Pharmacol* **2004**;4(4):372-7.
- [20.] Mavropoulos A, Orfanidou T, Liaskos C, Smyk DS, Billinis C, Blank M, Rigopoulou EIand Bogdanos DP. p38 mitogen-activated protein kinase (p38 MAPK)-mediated autoimmunity: lessons to learn from ANCA vasculitis and pemphigus vulgaris. *Autoimmunity reviews* **2013**;12(5):580-90.
- [21.] Mavropoulos A, Rigopoulou EI, Liaskos C, Bogdanos DPand Sakkas LI. The role of p38 MAPK in the aetiopathogenesis of psoriasis and psoriatic arthritis. *Clin Dev Immunol* **2013**;2013:569751.
- [22.] Mavropoulos A, Sully G, Cope APand Clark AR. Stabilization of IFN-gamma mRNA by MAPK p38 in IL-12- and IL-18-stimulated human NK cells. *Blood* **2005**;105(1):282-8.
- [23.] Arthur JSand Ley SC. Mitogen-activated protein kinases in innate immunity. *Nat Rev Immunol* **2013**;13(9):679-92.
- [24.] Sakkas LI, Bogdanos DP, Katsiari Cand Platsoucas CD. Anti-citrullinated peptides as autoantigens in rheumatoid arthritis-relevance to treatment. *Autoimmunity reviews* **2014**;13(11):1114-20.
- [25.] Han J, Lee JD, Bibbs Land Ulevitch RJ. A MAP kinase targeted by endotoxin and hyperosmolarity in mammalian cells. *Science* **1994**;265(5173):808-11.
- [26.] Lee JC, Laydon JT, McDonnell PC, Gallagher TF, Kumar S, Green D, McNulty D, Blumenthal MJ, Heys JR, Landvatter SWand et al. A protein kinase involved in the regulation of inflammatory cytokine biosynthesis. *Nature* **1994**;372(6508):739-46.
- [27.] Freshney NW, Rawlinson L, Guesdon F, Jones E, Cowley S, Hsuan Jand Saklatvala J. Interleukin-1 activates a novel protein kinase cascade that results in the phosphorylation of Hsp27. *Cell* **1994**;78(6):1039-49.
- [28.] Clark ARand Dean JL. The p38 MAPK Pathway in Rheumatoid Arthritis: A Sideways Look. *Open Rheumatol J* **2012**;6:209-219.
- [29.] Jiang Y, Chen C, Li Z, Guo W, Gegner JA, Lin Sand Han J. Characterization of the structure and function of a new mitogen-activated protein kinase (p38beta). *J Biol Chem* **1996**;271(30):17920-6.
- [30.] Li Z, Jiang Y, Ulevitch RJand Han J. The primary structure of p38 gamma: a new member of p38 group of MAP kinases. *Biochem Biophys Res Commun* **1996**;228(2):334-40.
- [31.] Hale KK, Trollinger D, RihaneK Mand Manthey CL. Differential expression and activation of p38 mitogen-activated protein kinase alpha, beta, gamma, and delta in inflammatory cell lineages. *J Immunol* **1999**;162(7):4246-52.
- [32.] Xing B, Bachstetter ADand Van Eldik LJ. Deficiency in p38beta MAPK fails to inhibit cytokine production or protect neurons against inflammatory insult in in vitro and in vivo mouse models. *PLoS One* **2013**;8(2):e56852.
- [33.] Wilson KP, Fitzgibbon MJ, Caron PR, Griffith JP, Chen W, McCaffrey PG, Chambers SPand Su MS. Crystal structure of p38 mitogen-activated protein kinase. *J Biol Chem* **1996**;271(44):27696-700.
- [34.] Bukhtiyarova M, Northrop K, Chai X, Casper D, Karpusas Mand Springman E. Improved expression, purification, and crystallization of p38alpha MAP kinase. *Protein Expr Purif* **2004**;37(1):154-61.
- [35.] Salvador JM, Mittelstadt PR, Guszczynski T, Copeland TD, Yamaguchi H, Appella E, Fornace AJ, Jr.and Ashwell JD. Alternative p38 activation pathway mediated by T cell receptor-proximal tyrosine kinases. *Nat Immunol* **2005**;6(4):390-5.
- [36.] Ge B, Gram H, Di Padova F, Huang B, New L, Ulevitch RJ, Luo Yand Han J. MAPKK-independent activation of p38alpha mediated by TAB1-dependent



- autophosphorylation of p38alpha. *Science* **2002**;295(5558):1291-4.
- [37.] Derijard B, Raingeaud J, Barrett T, Wu IH, Han J, Ulevitch RJ and Davis RJ. Independent human MAP-kinase signal transduction pathways defined by MEK and MKK isoforms. *Science* **1995**;267(5198):682-5.
- [38.] Dong C, Davis RJ and Flavell RA. MAP kinases in the immune response. *Annu Rev Immunol* **2002**;20:55-72.
- [39.] Cheung PC, Campbell DG, Nebreda A and Cohen P. Feedback control of the protein kinase TAK1 by SAPK2a/p38alpha. *EMBO J* **2003**;22(21):5793-805.
- [40.] Liebermann DA and Hoffman B. Gadd45 in stress signaling. *J Mol Signal* **2008**;3:15.
- [41.] Tan Y, Rouse J, Zhang A, Cariati S, Cohen P and Comb MJ. FGF and stress regulate CREB and ATF-1 via a pathway involving p38 MAP kinase and MAPKAP kinase-2. *EMBO J* **1996**;15(17):4629-42.
- [42.] Gaestel M. MAPKAP kinases - MKs - two's company, three's a crowd. *Nat Rev Mol Cell Biol* **2006**;7(2):120-30.
- [43.] Ananieva O, Darragh J, Johansen C, Carr JM, McIlrath J, Park JM, Wingate A, Monk CE, Toth R, Santos SG, Iversen L and Arthur JS. The kinases MSK1 and MSK2 act as negative regulators of Toll-like receptor signaling. *Nat Immunol* **2008**;9(9):1028-36.
- [44.] New L, Jiang Y, Zhao M, Liu K, Zhu W, Flood LJ, Kato Y, Parry GC and Han J. PRAK, a novel protein kinase regulated by the p38 MAP kinase. *EMBO J* **1998**;17(12):3372-84.
- [45.] Knight JD, Tian R, Lee RE, Wang F, Beauvais A, Zou H, Megeny LA, Gingras AC, Pawson T, Figeys D and Kothary R. A novel whole-cell lysate kinase assay identifies substrates of the p38 MAPK in differentiating myoblasts. *Skelet Muscle* **2012**;2:5.
- [46.] Risco A, del Fresno C, Mambol A, Alsina-Beauchamp D, MacKenzie KF, Yang HT, Barber DF, Morcelle C, Arthur JS, Ley SC, Ardavin C and Cuenda A. p38gamma and p38delta kinases regulate the Toll-like receptor 4 (TLR4)-induced cytokine production by controlling ERK1/2 protein kinase pathway activation. *Proc Natl Acad Sci U S A* **2012**;109(28):11200-5.
- [47.] Anderson P. Post-transcriptional control of cytokine production. *Nat Immunol* **2008**;9(4):353-9.
- [48.] Dean JL, Sully G, Clark AR and Saklatvala J. The involvement of AU-rich element-binding proteins in p38 mitogen-activated protein kinase pathway-mediated mRNA stabilisation. *Cell Signal* **2004**;16(10):1113-21.
- [49.] Clark A, Dean J, Tudor C and Saklatvala J. Post-transcriptional gene regulation by MAP kinases via AU-rich elements. *Front Biosci* **2009**;14:847-71.
- [50.] Khabar KS. Post-transcriptional control of cytokine gene expression in health and disease. *J Interferon Cytokine Res* **2014**;34(4):215-9.
- [51.] Brook M, Tchen CR, Santalucia T, McIlrath J, Arthur JS, Saklatvala J and Clark AR. Posttranslational regulation of tristetraprolin subcellular localization and protein stability by p38 mitogen-activated protein kinase and extracellular signal-regulated kinase pathways. *Mol Cell Biol* **2006**;26(6):2408-18.
- [52.] Chen P, Li J, Barnes J, Kokkonen GC, Lee JC and Liu Y. Restraint of proinflammatory cytokine biosynthesis by mitogen-activated protein kinase phosphatase-1 in lipopolysaccharide-stimulated macrophages. *J Immunol* **2002**;169(11):6408-16.
- [53.] Clark AR. MAP kinase phosphatase 1: a novel mediator of biological effects of glucocorticoids? *J Endocrinol* **2003**;178(1):5-12.
- [54.] Keyse SM. Dual-specificity MAP kinase phosphatases (MKPs) and cancer. *Cancer Metastasis Rev* **2008**;27(2):253-61.
- [55.] Tanoue T, Yamamoto T, Maeda R and Nishida E. A Novel MAPK phosphatase MKP-7 acts preferentially on JNK/SAPK and p38 alpha and beta MAPKs. *J Biol Chem* **2001**;276(28):26629-39.
- [56.] Adams RH, Porras A, Alonso G, Jones M, Vintersten K, Panelli S, Valladares A, Perez L, Klein R and Nebreda AR. Essential role of p38alpha MAP kinase in placental but not embryonic cardiovascular development. *Mol Cell* **2000**;6(1):109-16.
- [57.] Beardmore VA, Hinton HJ, Eftychi C, Apostolaki M, Armaka M, Darragh J, McIlrath J, Carr JM, Armit LJ, Clacher C, Malone L, Kollias G and Arthur JS. Generation and characterization of p38beta (MAPK11) gene-targeted mice. *Mol Cell Biol* **2005**;25(23):10454-64.
- [58.] Badger AM, Bradbeer JN, Votta B, Lee JC, Adams JL and Griswold DE. Pharmacological profile of SB 203580, a selective inhibitor of cytokine suppressive binding protein/p38 kinase, in animal models of arthritis, bone resorption, endotoxin shock and immune function. *J Pharmacol Exp Ther* **1996**;279(3):1453-61.
- [59.] Ridley SH, Dean JL, Sarsfield SJ, Brook M, Clark AR and Saklatvala J. A p38 MAP kinase inhibitor regulates stability of interleukin-1-induced cyclooxygenase-2 mRNA. *FEBS Lett* **1998**;439(1-2):75-80.
- [60.] Kumar S, Jiang MS, Adams JL and Lee JC. Pyridinylimidazole compound SB 203580 inhibits the activity but not the activation of p38 mitogen-activated protein kinase. *Biochem Biophys Res Commun* **1999**;263(3):825-31.
- [61.] Lee JC, Kassis S, Kumar S, Badger A and Adams JL. p38 mitogen-activated protein kinase inhibitors--mechanisms and therapeutic potentials. *Pharmacol Ther* **1999**;82(2-3):389-97.
- [62.] Foster ML, Halley F and Souness JE. Potential of p38 inhibitors in the treatment of rheumatoid arthritis. *Drug News Perspect* **2000**;13(8):488-97.
- [63.] Nemoto S, Xiang J, Huang S and Lin A. Induction of apoptosis by SB202190 through inhibition of p38beta mitogen-activated protein kinase. *J Biol Chem* **1998**;273(26):16415-20.
- [64.] Schett G, Zwerina J and Firestein G. The p38 mitogen-activated protein kinase (MAPK) pathway in rheumatoid arthritis. *Ann Rheum Dis* **2008**;67(7):909-16.
- [65.] Sweeney SE. The as-yet unfulfilled promise of p38 MAPK inhibitors. *Nat Rev Rheumatol* **2009**;5(9):475-7.



- [66.] Hill RJ, Dabbagh K, Phippard D, Li C, Suttman RT, Welch M, Papp E, Song KW, Chang KC, Leaffer D, Kim YN, Roberts RT, Zabka TS, Aud D, Dal Porto J, Manning AM, Peng SL, Goldstein DM and Wong BR. Pamapimod, a novel p38 mitogen-activated protein kinase inhibitor: preclinical analysis of efficacy and selectivity. *J Pharmacol Exp Ther* **2008**;327(3):610-9.
- [67.] Kyttaris VC. Kinase inhibitors: a new class of antirheumatic drugs. *Drug Des Devel Ther* **2012**;6:245-50.
- [68.] Buhler Sand Laufer SA. p38 MAPK inhibitors: a patent review (2012 - 2013). *Expert Opin Ther Pat* **2014**;24(5):535-54.
- [69.] Haase I, Hobbs RM, Romero MR, Broad Sand Watt FM. A role for mitogen-activated protein kinase activation by integrins in the pathogenesis of psoriasis. *J Clin Invest* **2001**;108(4):527-36.
- [70.] Johansen C, Kragballe K, Westergaard M, Henningsen J, Kristiansen Kand Iversen L. The mitogen-activated protein kinases p38 and ERK1/2 are increased in lesional psoriatic skin. *Br J Dermatol* **2005**;152(1):37-42.
- [71.] Lang R, Hammer Mand Mages J. DUSP meet immunology: dual specificity MAPK phosphatases in control of the inflammatory response. *J Immunol* **2006**;177(11):7497-504.
- [72.] Kjellerup RB, Johansen C, Kragballe Kand Iversen L. The expression of dual specificity phosphatase 1 mRNA is downregulated in lesional psoriatic skin. *Br J Dermatol* **2012**.
- [73.] Johansen C, Funding AT, Otkjaer K, Kragballe K, Jensen UB, Madsen M, Binderup L, Skak-Nielsen T, Fjording MS and Iversen L. Protein expression of TNF-alpha in psoriatic skin is regulated at a posttranscriptional level by MAPK-activated protein kinase 2. *J Immunol* **2006**;176(3):1431-8.
- [74.] Kotlyarov A, Neininger A, Schubert C, Eckert R, Birchmeier C, Volk HD and Gaestel M. MAPKAP kinase 2 is essential for LPS-induced TNF-alpha biosynthesis. *Nat Cell Biol* **1999**;1(2):94-7.
- [75.] Hegen M, Gaestel M, Nickerson-Nutter CL, Lin L and Telliez JB. MAPKAP kinase 2-deficient mice are resistant to collagen-induced arthritis. *J Immunol* **2006**;177(3):1913-7.
- [76.] Duraisamy S, Bajpai M, Bughani U, Dastidar SG, Ray A and Chopra P. MK2: a novel molecular target for anti-inflammatory therapy. *Expert Opin Ther Targets* **2008**;12(8):921-36.
- [77.] Gaestel M, Kotlyarov A and Kracht M. Targeting innate immunity protein kinase signalling in inflammation. *Nat Rev Drug Discov* **2009**;8(6):480-99.
- [78.] Funding AT, Johansen C, Kragballe Kand Iversen L. Mitogen- and stress-activated protein kinase 2 and cyclic AMP response element binding protein are activated in lesional psoriatic epidermis. *J Invest Dermatol* **2007**;127(8):2012-9.
- [79.] Soegaard-Madsen L, Johansen C, Iversen Land Kragballe K. Adalimumab therapy rapidly inhibits p38 mitogen-activated protein kinase activity in lesional psoriatic skin preceding clinical improvement. *Br J Dermatol* **2010**;162(6):1216-23.
- [80.] Gesser B, Johansen C, Rasmussen MK, Funding AT, Otkjaer K, Kjellerup RB, Kragballe Kand Iversen L. Dimethylfumarate specifically inhibits the mitogen and stress-activated kinases 1 and 2 (MSK1/2): possible role for its anti-psoriatic effect. *J Invest Dermatol* **2007**;127(9):2129-37.
- [81.] Yu XJ, Li CY, Dai HY, Cai DX, Wang KY, Xu YH, Chen L and Zhou CL. Expression and localization of the activated mitogen-activated protein kinase in lesional psoriatic skin. *Exp Mol Pathol* **2007**;83(3):413-8.
- [82.] Ikewaki N and Inoko H. A very late activating antigen-alpha4 (CD49d) monoclonal antibody, BU49 induces phosphorylation of a cAMP response element-binding protein (CREB), resulting in induction of homotypic cell aggregation and enhancement of interleukin-8 (IL-8) production. *Microbiol Immunol* **2002**;46(10):685-95.
- [83.] Dauletbaev N, Eklove D, Mawji N, Iskandar M, Di Marco S, Gallouzi IE and Lands LC. Down-regulation of cytokine-induced interleukin-8 requires inhibition of p38 mitogen-activated protein kinase (MAPK) via MAPK phosphatase 1-dependent and -independent mechanisms. *J Biol Chem* **2011**;286(18):15998-6007.
- [84.] Johansen C, Vinter H, Soegaard-Madsen L, Olsen LR, Steiniche T, Iversen Land Kragballe K. Preferential inhibition of the mRNA expression of p38 mitogen-activated protein kinase regulated cytokines in psoriatic skin by anti-TNFalpha therapy. *Br J Dermatol* **2010**;163(6):1194-204.
- [85.] Andres RM, Hald A, Johansen C, Kragballe Kand Iversen L. Studies of Jak/STAT3 expression and signalling in psoriasis identifies STAT3-Ser727 phosphorylation as a modulator of transcriptional activity. *Exp Dermatol* **2013**;22(5):323-8.
- [86.] Hald A, Andres RM, Salskov-Iversen ML, Kjellerup RB, Iversen Land Johansen C. STAT1 expression and activation is increased in lesional psoriatic skin. *Br J Dermatol* **2013**;168(2):302-10.
- [87.] Sano S, Chan KS, Carbajal S, Clifford J, Peavey M, Kiguchi K, Itami S, Nickoloff BJ and DiGiovanni J. Stat3 links activated keratinocytes and immunocytes required for development of psoriasis in a novel transgenic mouse model. *Nat Med* **2005**;11(1):43-9.
- [88.] Yego EC and Dillman JF, 3rd. Cytokine regulation by MAPK activated kinase 2 in keratinocytes exposed to sulfur mustard. *Toxicol In Vitro* **2013**;27(7):2067-75.
- [89.] Sherlock JP, Taylor PC and Buckley CD. The biology of IL-23 and IL-17 and their therapeutic targeting in rheumatic diseases. *Curr Opin Rheumatol* **2015**;27(1):71-5.
- [90.] Mease PJ. Inhibition of interleukin-17, interleukin-23 and the TH17 cell pathway in the treatment of psoriatic arthritis and psoriasis. *Curr Opin Rheumatol* **2015**;27(2):127-33.
- [91.] Said A, Bock S, Lajqi T, Muller Gand Weindl G. Chloroquine promotes IL-17 production by CD4+ T cells



- via p38-dependent IL-23 release by monocyte-derived Langerhans-like cells. *J Immunol* **2014**;193(12):6135-43.
- [92.] Peral de Castro C, Jones SA, Ni Cheallaigh C, Hearnden CA, Williams L, Winter J, Lavelle EC, Mills KHand Harris J. Autophagy regulates IL-23 secretion and innate T cell responses through effects on IL-1 secretion. *J Immunol* **2012**;189(8):4144-53.
- [93.] Diani M, Altomare Gand Reali E. T cell responses in psoriasis and psoriatic arthritis. *Autoimmunity reviews* **2014**.
- [94.] Schaper K, Kietzmann Mand Baumer W. Sphingosine-1-phosphate differently regulates the cytokine production of IL-12, IL-23 and IL-27 in activated murine bone marrow derived dendritic cells. *Mol Immunol* **2014**;59(1):10-8.
- [95.] Mose M, Kang Z, Raaby L, Iversen Land Johansen C. TNFalpha- and IL-17A-mediated S100A8 expression is regulated by p38 MAPK. *Exp Dermatol* **2013**;22(7):476-81.
- [96.] Hau CS, Kanda N, Noda S, Tatsuta A, Kamata M, Shibata S, Asano Y, Sato S, Watanabe Sand Tada Y. Visfatin enhances the production of cathelicidin antimicrobial peptide, human beta-defensin-2, human beta-defensin-3, and S100A7 in human keratinocytes and their orthologs in murine imiquimod-induced psoriatic skin. *Am J Pathol* **2013**;182(5):1705-17.
- [97.] Yamamoto M, Nakajima K, Takaishi M, Kitaba S, Magata Y, Kataoka Sand Sano S. Psoriatic inflammation facilitates the onset of arthritis in a mouse model. *J Invest Dermatol* **2015**;135(2):445-53.
- [98.] Cauli Aand Mathieu A. Th17 and interleukin 23 in the pathogenesis of psoriatic arthritis and spondyloarthritis. *J Rheumatol Suppl* **2012**;89:15-8.
- [99.] Celis R, Planell N, Fernandez-Sueiro JL, Sanmarti R, Ramirez J, Gonzalez-Alvaro I, Pablos JLand Canete JD. Synovial cytokine expression in psoriatic arthritis and associations with lymphoid neogenesis and clinical features. *Arthritis Res Ther* **2012**;14(2):R93.
- [100.] Candia L, Marquez J, Hernandez C, Zea AHand Espinoza LR. Toll-like receptor-2 expression is upregulated in antigen-presenting cells from patients with psoriatic arthritis: a pathogenic role for innate immunity? *J Rheumatol* **2007**;34(2):374-9.
- [101.] Amital H, Barak V, Winkler REand Rubinow A. Impact of treatment with infliximab on serum cytokine profile of patients with rheumatoid and psoriatic arthritis. *Ann N Y Acad Sci* **2007**;1110:649-60.
- [102.] Cordiali-Fei P, Ardigo M, Mastroianni A, Giuliani A, G DA, Bordignon V, Trento E, Vento Aand Berardesca E. Serum cytokines and biohumoral immunological characterization of psoriatic patients in long term etanercept treatment. *Int J Immunopathol Pharmacol* **2008**;21(3):643-9.
- [103.] de Vlam Kand Lories RJ. Efficacy, effectiveness and safety of etanercept in monotherapy for refractory psoriatic arthritis: a 26-week observational study. *Rheumatology (Oxford)* **2006**;45(3):321-4.
- [104.] Danning CL, Illei GG, Hitchon C, Greer MR, Boumpas DTand McInnes IB. Macrophage-derived cytokine and nuclear factor kappaB p65 expression in synovial membrane and skin of patients with psoriatic arthritis. *Arthritis Rheum* **2000**;43(6):1244-56.
- [105.] Lories RJ, Derese I, Luyten FPand de Vlam K. Activation of nuclear factor kappa B and mitogen activated protein kinases in psoriatic arthritis before and after etanercept treatment. *Clin Exp Rheumatol* **2008**;26(1):96-102.
- [106.] Frey S, Derer A, Messbacher ME, Baeten DL, Bugatti S, Montecucco C, Schett Gand Hueber AJ. The novel cytokine interleukin-36alpha is expressed in psoriatic and rheumatoid arthritis synovium. *Ann Rheum Dis* **2013**.
- [107.] Vervoordeldonk MJand Tak PP. Cytokines in rheumatoid arthritis. *Curr Rheumatol Rep* **2002**;4(3):208-17.
- [108.] Fournier C. Where do T cells stand in rheumatoid arthritis? *Joint Bone Spine* **2005**;72(6):527-32.
- [109.] Okamoto H, Hoshi D, Kiire A, Yamanaka Hand Kamatani N. Molecular targets of rheumatoid arthritis. *Inflamm Allergy Drug Targets* **2008**;7(1):53-66.
- [110.] Aita T, Yamamura M, Kawashima M, Okamoto A, Iwahashi M, Yamana Jand Makino H. Expression of interleukin 12 receptor (IL-12R) and IL-18R on CD4+ T cells from patients with rheumatoid arthritis. *J Rheumatol* **2004**;31(3):448-56.
- [111.] Gracie JA, Forsey RJ, Chan WL, Gilmour A, Leung BP, Greer MR, Kennedy K, Carter R, Wei XQ, Xu D, Field M, Foulis A, Liew FYand McInnes IB. A proinflammatory role for IL-18 in rheumatoid arthritis. *J Clin Invest* **1999**;104(10):1393-401.
- [112.] Dodeller F, Skapenko A, Kalden JR, Lipsky PEand Schulze-Koops H. The p38 mitogen-activated protein kinase regulates effector functions of primary human CD4 T cells. *Eur J Immunol* **2005**;35(12):3631-42.
- [113.] Brennan FM, Hayes AL, Ciesielski CJ, Green P, Foxwell BManand Feldmann M. Evidence that rheumatoid arthritis synovial T cells are similar to cytokine-activated T cells: involvement of phosphatidylinositol 3-kinase and nuclear factor kappaB pathways in tumor necrosis factor alpha production in rheumatoid arthritis. *Arthritis Rheum* **2002**;46(1):31-41.
- [114.] Li C, Beavis P, Palfreeman AC, Amjadi P, Kennedy Aand Brennan FM. Activation of p38 mitogen-activated protein kinase is critical step for acquisition of effector function in cytokine-activated T cells, but acts as a negative regulator in T cells activated through the T-cell receptor. *Immunology* **2011**;132(1):104-10.
- [115.] Campbell J, Ciesielski CJ, Hunt AE, Horwood NJ, Beech JT, Hayes LA, Denys A, Feldmann M, Brennan FMand Foxwell BM. A novel mechanism for TNF-alpha regulation by p38 MAPK: involvement of NF-kappa B with implications for therapy in rheumatoid arthritis. *J Immunol* **2004**;173(11):6928-37.
- [116.] Tang C, Li Y, Lin X, Ye J, Li W, He Z, Li Fand Cai X. Prolactin increases tumor necrosis factor alpha expression in peripheral CD14 monocytes of patients



- with rheumatoid arthritis. *Cell Immunol* **2014**;290(1):164-8.
- [117.] Roeleveld DMand Koenders MI. The role of the Th17 cytokines IL-17 and IL-22 in Rheumatoid Arthritis pathogenesis and developments in cytokine immunotherapy. *Cytokine* **2014**.
- [118.] Dong Wand Zhu P. Functional niche of inflamed synovium for Th17-cell expansion and activation in rheumatoid arthritis: implication to clinical therapeutics. *Autoimmunity reviews* **2012**;11(12):844-51.
- [119.] Wei R, Dong L, Xiao Q, Sun D, Li Xand Nian H. Engagement of Toll-like receptor 2 enhances interleukin (IL)-17(+) autoreactive T cell responses via p38 mitogen-activated protein kinase signalling in dendritic cells. *Clin Exp Immunol* **2014**;178(2):353-63.
- [120.] Kleinewietfeld M, Manzel A, Titze J, Kvakana H, Yosef N, Linker RA, Muller DNand Hafler DA. Sodium chloride drives autoimmune disease by the induction of pathogenic TH17 cells. *Nature* **2013**;496(7446):518-22.
- [121.] Dulos J, Wijnands FP, van den Hurk-van Alebeek JA, van Vugt MJ, Rullmann JA, Schot JJ, de Groot MW, Wagenaars JL, van Ravestein-van Os R, Smets RL, Vink PM, Hofstra CL, Nelissen RLand van Eenennaam H. p38 inhibition and not MK2 inhibition enhances the secretion of chemokines from TNF-alpha activated rheumatoid arthritis fibroblast-like synoviocytes. *Clin Exp Rheumatol* **2013**;31(4):515-25.
- [122.] Inoue T, Hammaker D, Boyle DLand Firestein GS. Regulation of p38 MAPK by MAPK kinases 3 and 6 in fibroblast-like synoviocytes. *J Immunol* **2005**;174(7):4301-6.
- [123.] Chabaud-Riou Mand Firestein GS. Expression and activation of mitogen-activated protein kinase kinases-3 and -6 in rheumatoid arthritis. *Am J Pathol* **2004**;164(1):177-84.
- [124.] Burnette BL, Selness S, Devraj R, Jungbluth G, Kurumbail R, Stillwell L, Anderson G, Mnich S, Hirsch J, Compton R, De Ciechi P, Hope H, Hepperle M, Keith RH, Naing W, Shieh H, Portanova J, Zhang Y, Zhang J, Leimgruber RMand Monahan J. SD0006: a potent, selective and orally available inhibitor of p38 kinase. *Pharmacology* **2009**;84(1):42-60.
- [125.] Revesz L, Blum E, Di Padova FE, Buhl T, Feifel R, Gram H, Hiestand P, Manning Uand Rucklin G. Novel p38 inhibitors with potent oral efficacy in several models of rheumatoid arthritis. *Bioorg Med Chem Lett* **2004**;14(13):3595-9.
- [126.] Mihara K, Almansa C, Smeets RL, Loomans EE, Dulos J, Vink PM, Rooseboom M, Kreutzer H, Cavalcanti F, Boots AMand Nelissen RL. A potent and selective p38 inhibitor protects against bone damage in murine collagen-induced arthritis: a comparison with neutralization of mouse TNFalpha. *Br J Pharmacol* **2008**;154(1):153-64.
- [127.] Guma M, Hammaker D, Topolewski K, Corr M, Boyle DL, Karin Mand Firestein GS. Antiinflammatory functions of p38 in mouse models of rheumatoid arthritis: advantages of targeting upstream kinases MKK-3 or MKK-6. *Arthritis Rheum* **2012**;64(9):2887-95.
- [128.] Dambach DM. Potential adverse effects associated with inhibition of p38alpha/beta MAP kinases. *Curr Top Med Chem* **2005**;5(10):929-39.
- [129.] Damjanov N, Kauffman RSand Spencer-Green GT. Efficacy, pharmacodynamics, and safety of VX-702, a novel p38 MAPK inhibitor, in rheumatoid arthritis: results of two randomized, double-blind, placebo-controlled clinical studies. *Arthritis Rheum* **2009**;60(5):1232-41.
- [130.] Terajima M, Inoue T, Magari K, Yamazaki H, Higashi Yand Mizuhara H. Anti-inflammatory effect and selectivity profile of AS1940477, a novel and potent p38 mitogen-activated protein kinase inhibitor. *Eur J Pharmacol* **2013**;698(1-3):455-62.
- [131.] Montalban AG, Boman E, Chang CD, Ceide SC, Dahl R, Dalesandro D, Delaet NG, Erb E, Ernst J, Gibbs A, Kahl J, Kessler L, Lundstrom J, Miller S, Nakanishi H, Roberts E, Saiah E, Sullivan R, Wang Zand Larson CJ. KR-003048, a potent, orally active inhibitor of p38 mitogen-activated protein kinase. *Eur J Pharmacol* **2010**;632(1-3):93-102.
- [132.] Hammaker D, Boyle DL, Topolewski Kand Firestein GS. Differential regulation of anti-inflammatory genes by p38 MAP kinase and MAP kinase kinase 6. *J Inflamm (Lond)* **2014**;11:14.
- [133.] Ellis SD, McGovern JL, van Maurik A, Howe D, Ehrenstein MRand Notley CA. Induced CD8+FoxP3+ Treg cells in rheumatoid arthritis are modulated by p38 phosphorylation and monocytes expressing membrane tumor necrosis factor alpha and CD86. *Arthritis Rheumatol* **2014**;66(10):2694-705.
- [134.] Adler HS, Kubsch S, Graulich E, Ludwig S, Knop Jand Steinbrink K. Activation of MAP kinase p38 is critical for the cell-cycle-controlled suppressor function of regulatory T cells. *Blood* **2007**;109(10):4351-9.
- [135.] Adler HSand Steinbrink K. MAP kinase p38 and its relation to T cell anergy and suppressor function of regulatory T cells. *Cell Cycle* **2008**;7(2):169-70.
- [136.] Ohkusu-Tsukada K, Toda M, Udono H, Kawakami Yand Takahashi K. Targeted inhibition of IL-10-secreting CD25- Treg via p38 MAPK suppression in cancer immunotherapy. *Eur J Immunol*;40(4):1011-21.
- [137.] Koczan D, Guthke R, Thiesen HJ, Ibrahim SM, Kundt G, Krentz H, Gross Gand Kunz M. Gene expression profiling of peripheral blood mononuclear leukocytes from psoriasis patients identifies new immune regulatory molecules. *Eur J Dermatol* **2005**;15(4):251-7.
- [138.] Krutzik PO, Irish JM, Nolan GPand Perez OD. Analysis of protein phosphorylation and cellular signaling events by flow cytometry: techniques and clinical applications. *Clin Immunol* **2004**;110(3):206-21.
- [139.] Aerts NE, Ebo DG, Bridts CH, Stevens WJand De Clerck LS. Flow cytometric analysis of phospho-p38 mitogen-activated kinase (MAPK): p38 MAPK does not mediate the effect of adalimumab on peripheral T cell cytokine production in rheumatoid arthritis. *Cytokine* **2009**;47(3):178-84.
- [140.] Galligan CL, Siebert JC, Siminovitich KA, Keystone EC, Bykerk V, Perez ODand Fish EN.

Multiparameter phospho-flow analysis of lymphocytes in early rheumatoid arthritis: implications for diagnosis and monitoring drug therapy. *PLoS One* **2009**;4(8):e6703.

[141.] Mavropoulos A, Smyk D, Rigopoulou EI and Bogdanos DP. Human peripheral blood mononuclear cell culture for flow cytometric analysis of phosphorylated mitogen-activated protein kinases. *Methods Mol Biol* **2012**;806:275-85.

[142.] Mavropoulos A, Bogdanos DP, Liaskos C, Orfanidou T, Simopoulou T, Zafiriou E, Sakkas LI and Rigopoulou EI. Flow cytometric detection of p38 MAPK phosphorylation and intracellular cytokine expression in peripheral blood subpopulations from patients with autoimmune rheumatic diseases. *J Immunol Res* **2014**;2014:671431.

[143.] Abecassis S, Giustiniani J, Meyer N, Schiavon V, Ortonne N, Campillo JA, Bagot Mand Bensussan A. Identification of a novel CD160+ CD4+ T-lymphocyte subset in the skin: a possible role for CD160 in skin inflammation. *J Invest Dermatol* **2007**;127(5):1161-6.

[144.] Batliwalla FM, Li W, Ritchlin CT, Xiao X, Brenner M, Laragione T, Shao T, Durham R, Kemshetti S, Schwarz E, Coe R, Kern M, Baechler EC, Behrens TW, Gregersen PK and Gulko PS. Microarray analyses of peripheral blood cells identifies unique gene expression signature in psoriatic arthritis. *Mol Med* **2005**;11(1-12):21-9.

[145.] Bowcock AM, Shannon W, Du F, Duncan J, Cao K, Aftergut K, Catier J, Fernandez-Vina MA and Menter A. Insights into psoriasis and other inflammatory diseases from large-scale gene expression studies. *Hum Mol Genet* **2001**;10(17):1793-805.

[146.] Quekenborn-Trinquet V, Fogel P, Aldana-Jammayrac O, Ancian P, Demarchez M, Rossio P, Richards HL, Kirby B, Nguyen C, Voegel JJ and Griffiths CE. Gene expression profiles in psoriasis: analysis of impact of body site location and clinical severity. *Br J Dermatol* **2005**;152(3):489-504.

[147.] Mavropoulos A, Spyrou E, Rigopoulou EI, Vergani D, Dalekos GN and Bogdanos DP. 1107 Phosphorylation of P38 MAPK is detectable in NKT cells of patients with autoimmune hepatitis in whom it mirrors disease activity. *Journal of hepatology*;52:S428.



Dr A. Mavropoulos holds a BSc degree in Genetics, an MSc degree in Medical Molecular Genetics and a PhD in Immunology awarded from the Kennedy Institute of Rheumatology Division, Imperial College of Science Technology and Medicine, University of London, UK. He is currently a senior post-doctoral fellow in Cellular Immunotherapy and Molecular Immunodiagnosics, Institute of Research and Technology of Thessaly, Larissa, Greece and also affiliated with the Department of Rheumatology, Faculty of Medicine, School of Health Sciences, University of Thessaly, Greece

Dr Mavropoulos has been awarded the prestigious 2008 Sheila Sherlock Post-Doctoral Fellowship from the European Association for the Study of the Liver for his work on phospho-flow cytometric determination of p38 MAPK signalling cascade in autoimmune liver diseases. In 2005, published data in *Blood Journal* demonstrating an innovative mechanism of IFN- γ mRNA stabilization mediated by the p38 MAPK pathway in NK cells stimulated by IL-12 and IL-18.



Dimitrios P. Bogdanos is an affiliated member of the Department of Rheumatology, Faculty of Medicine, School of Health Sciences, Larissa, Greece and is also heading the Cellular Immunotherapy and Molecular Immunodiagnosics Group at the Institute of Research and Technology of Thessaly, Larissa, Greece. He has been awarded an MD from the Aristotle University of Thessaloniki (1997) and a PhD from the University College London, University of London (2005). He has



been focused on the molecular mimicry project at King's College London (2003-2011) and awarded a CLS Award from HEFCE (2007). He has obtained a Professorship in Liver Immunopathology (Hon), Division of Transplantation Immunology and Mucosal Biology, King's College London (2011). He has earned more than 15 prizes-awards for his scientific work on the immunopathogenesis of microbial-triggered, including the Dame Sheila Sherlock Award from the British Association for the Study of the Liver (2005). He is an editor, associate editor or member of the editorial board in more than 15 journals.

SUBMITTING YOUR PAPER FOR REVIEW

A. Conditions to publish

It is a condition of publication that manuscripts submitted to this journal have not been published and will not be simultaneously submitted or published elsewhere. Plagiarism is strictly forbidden, and by submitting the article for publication the authors agree that the publishers have the legal right to take appropriate action against the authors, if plagiarism or fabricated information is discovered. By submitting a manuscript the authors agree that the copyright of their article is transferred to the publishers if and when the article is accepted for publication. Once submitted to the journal, the authors will not withdraw their manuscript at any stage prior to publication move elsewhere

B. Review Stage Using Word 6.0 or Higher

If you want to submit your file with one column electronically, please do the following:

--First, click on the View menu and choose Print Layout.

--Second, place your cursor in the first paragraph. Go to the Format menu, choose Columns, choose one column Layout, and choose "apply to whole document" from the dropdown menu.

--Third, click and drag the right margin bar to just over 4 inches in width.

The graphics will stay in the "second" column, but you can drag them to the first column. Make the graphic wider to push out any text that may try to fill in next to the graphic.

C. Final Stage Using Word 6.0

When you submit your final version (after your paper has been accepted), print it in two-column format, including figures and tables. You could send your final manuscript via e-mail or through a Web manuscript submission system as directed by the society contact. You may use *Zip* for large files, or compress files using *Stuffit*, *Compress*, *Pkzip*, or *Gzip*.

Also, send a sheet of paper or PDF with complete contact information for all authors. Include full mailing addresses, telephone numbers, fax numbers, and e-mail addresses. This

information will be used to send each author a complimentary copy of the journal in which the paper appears. In addition, designate one author as the "corresponding author." This is the author to whom proofs of the paper will be sent. Proofs are sent to the corresponding author only.

EDITORIAL POLICY

The submitting author is responsible for obtaining agreement of all coauthors and any consent required from sponsors before submitting a paper. We strongly discourages courtesy authorship. It is the obligation of the authors to cite relevant prior work.

PUBLICATION PRINCIPLES

Authors should consider the following points:

- 1) Technical papers submitted for publication must advance the state of knowledge and must cite relevant prior work.
- 2) To publish in our Journals, the Authors have to be sure to follow the rules concerning ethical publishing.
- 3) The length of a submitted paper should be commensurate with the importance, or appropriate to the complexity, of the work. For example, an obvious extension of previously published work might not be appropriate for publication or might be adequately treated in just a few pages.
- 4) Authors must convince peer reviewers and the editors of the scientific and technical merit of a paper; the standards of proof are higher when extraordinary or unexpected results are reported.
- 5) Because replication is required for scientific progress, papers submitted for publication must provide sufficient information to allow readers to perform similar experiments or calculations and use the reported results. Although not everything need be disclosed, a paper must contain new, useable, and fully described information. For example, a specimen's chemical composition need not be reported if the main purpose of a paper is to introduce a new measurement technique. Authors should expect to be challenged by reviewers if the results are not supported by adequate data and critical details.