

Overexpression of SMAD7 protects liver from TGFb/Smad-mediated fibrogenesis

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Background

Smad7 is a dominant intracellular inhibitor of TGFb/Activin signal transduction.¹ Recent animal model studies have corroborated the protective function of Smad7 in attenuating TGFb-mediated fibrosis in multiple organs, including liver, through manipulation of Smad7 expression.²⁻⁵

This study was scheduled in order to determine whether *Smad7* mRNA expression correlates with the expression of the molecules participating in the TGFb/Activin signal transduction pathway in liver tissue of patients with chronic hepatic diseases and to seek correlations with the status of liver inflammation, fibrosis and the effect of treatment.

Methods

Liver biopsies obtained from 67 patients with chronic hepatic diseases including a) 18 with chronic HCV hepatitis (CHC); b) 19 with chronic HBV hepatitis at diagnosis (CHB/d); c) 4 with CHB after antiviral treatment and relapse (CHB/nr) d) 14 with CHB after antiviral treatment response and remission for >5y (CHB/r); e) 12 with non alcoholic fatty liver disease (NAFLD). Three individuals submitted to liver biopsy due to a mild increase of aminotransferases but without liver architecture changes (served as controls). Demographic, clinicopathological and serological data of the analyzed subjects are summarized in **Table 1**.

The mRNA levels of *TGFb*s (*TGFb-1,-2,-3*), *activins* (*A,B,C,E*), *ALK4*, *ALK5*, *SMAD* molecules (*SMAD-2,-3,-4,-7*), and *CTGF* were determined in a quantitative reverse transcriptase PCR using SYBR-Green PCR Supermix (Invitrogen,UK). Primers were either designed or commercially obtained by SA Biosciences (USA). The sequences of the designed primers as well as the thermocycling conditions for all genes are summarized in **Table 2**. The *beta-2-microglobulin* (*B2M*) gene was used as a reference gene for sample normalization. Statistical analyses were performed using the SPSS ver. 18.0 software.

Table 1. Clinicopathological and serological data of the patients of the study

	Controls	CHB/d ^a	CHB/nr ^b	CHB/r ^c	CHC ^d	NAFLD ^e
No	3	19	4	14	18	12
Sex (M/F) ^f	2/1	9/10	2/2	11/3	14/4	7/5
Age (median, range)	61, 60-67	54, 24-64	57, 22-65	52, 23-60	41.5, 27-54	45, 21-71
AST ^g (U/ μ L), (median, range)	42, 36-45	51, 17-1969	62, 39-277	29.5, 15-51	45, 24-237	31.5, 19-70
ALT ^h (U/ μ L), (median, range)	32, 21-48	61, 15-1478	97.5, 70-332	31.5, 17-49	75, 32-213	54, 15-141
Inflammation grade ⁱ						
I-0 ⁱ	3	—	—	1	—	3
I-1 ⁱ	—	4	—	10	2	4
I-2 ⁱ	—	8	3	3	10	5
I-3 ⁱ	—	5	1	—	6	—
I-4 ⁱ	—	2	—	—	—	—
Fibrosis (median, range) ^j	—	4.0, 0-6	4.5, 1-5	2.0, 0-3	3.0, 1-6	0.5, 0-2
HAI-score (median, range)	—	8.0, 1-15	8.0, 5-11	2.0, 0-7	7.0, 2-12	2.0, 0-5
Viral load (median, range)	—	4 Meq/mL (0.009-699)	0.10 Meq/mL (0-44.5)	0 Meq/mL (0-0.008)	0.70 Meq/mL (0.10-6.25)	0, 0-0

Abbreviations: ^a CHB/d, newly diagnosed patients with Chronic HBV hepatitis; ^b CHB/nr, CHB patients 6 months after treatment withdrawal and no virologic/biochemical sustained response; ^c CHB/r, CHB patients after antiviral treatment response and remission for >5y; ^d CHC, Chronic HCV hepatitis; ^e NAFLD, non-alcoholic fatty liver disease; ^f M, male; F, female; ^g AST, aspartate aminotransferase; ^h ALT, alanine aminotransferase; ⁱ Inflammation grade (I-0: without inflammation, I-1: minimal, I-2: mild, I-3: moderate and I-4: marked) and fibrosis

Table 2. Primers and PCR conditions for the amplification of the analyzed genes

Gene	Primers	Sequence	PCR conditions
<i>TGFb1</i>	forward	commercially obtained by SABiosciences, Cat No PPH00508A	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
	reverse	commercially obtained by SABiosciences, Cat No PPH00531E	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 53°C for 15 s, 72°C for 15 s)
<i>TGFb2</i>	forward	5'- AgAgTgCCTgAACAA -3'	95°C for 2 min, followed by 40 cycles (95°C for 15 s, 53°C for 15 s, 72°C for 15 s)
	reverse	5'- CCATTGcCCTTCTCTCT -3'	95°C for 10 min, followed by 40 cycles (95°C for 10 s, 58°C for 10 s, 72°C for 30 s)
<i>TGFb3</i>	forward	commercially obtained by SABiosciences, Cat No PPH00531E	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
	reverse	commercially obtained by SABiosciences, Cat No PPH00237B	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
<i>ALK5</i>	forward	5'- CATTgACATTgCCCCgAATC -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
	reverse	5'- CgAgCAATCTCCCAATACAAg -3'	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 58°C for 15 s, 72°C for 15 s)
<i>SMAD2</i>	forward	commercially obtained by SABiosciences, Cat No PPH01949E	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 58°C for 10 s, 72°C for 30 s)
	reverse	commercially obtained by SABiosciences, Cat No PPH01921B	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
<i>SMAD3</i>	forward	commercially obtained by SABiosciences, Cat No PPH01921B	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
	reverse	commercially obtained by SABiosciences, Cat No PPH0134B	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
<i>SMAD4</i>	forward	commercially obtained by SABiosciences, Cat No PPH0134B	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
	reverse	commercially obtained by SABiosciences, Cat No PPH01905B	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
<i>CTGF</i>	forward	5'- ACCAATgACAAgCCTCCTg -3'	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
	reverse	5'- TTgCCTTCTTA ATgTTCTCTCC -3'	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
<i>INHBA</i>	forward	5'- AgCgACCTCgAgATCATC -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
	reverse	5'- TTgggGACTTTTAggAAgAgC -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
<i>INHBB</i>	forward	5'- AggAgCgCgTTTCCgAAATC -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
	reverse	5'- TggTTgCCTTgTggAgATg -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
<i>INHBC</i>	forward	5'- AgAgCgCgTTTgAgACTgC -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
	reverse	5'- AAgAgCgAgCTgTgTgATgTg -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
<i>INHBE</i>	forward	5'- gCAACAATTCCTgCgATACC -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
	reverse	5'- gCCTCAATTCCTCCAC -3'	95°C for 2 min, followed by 50 cycles (95°C for 15 s, 56°C for 40 s), and 72°C for 2 min
<i>B2M</i>	forward	commercially obtained by SABiosciences, Cat No PPH01094E	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)
	reverse	commercially obtained by SABiosciences, Cat No PPH01094E	95°C for 10 min, followed by 40 cycles (95°C for 15 s, 60°C for 60 s)

Results

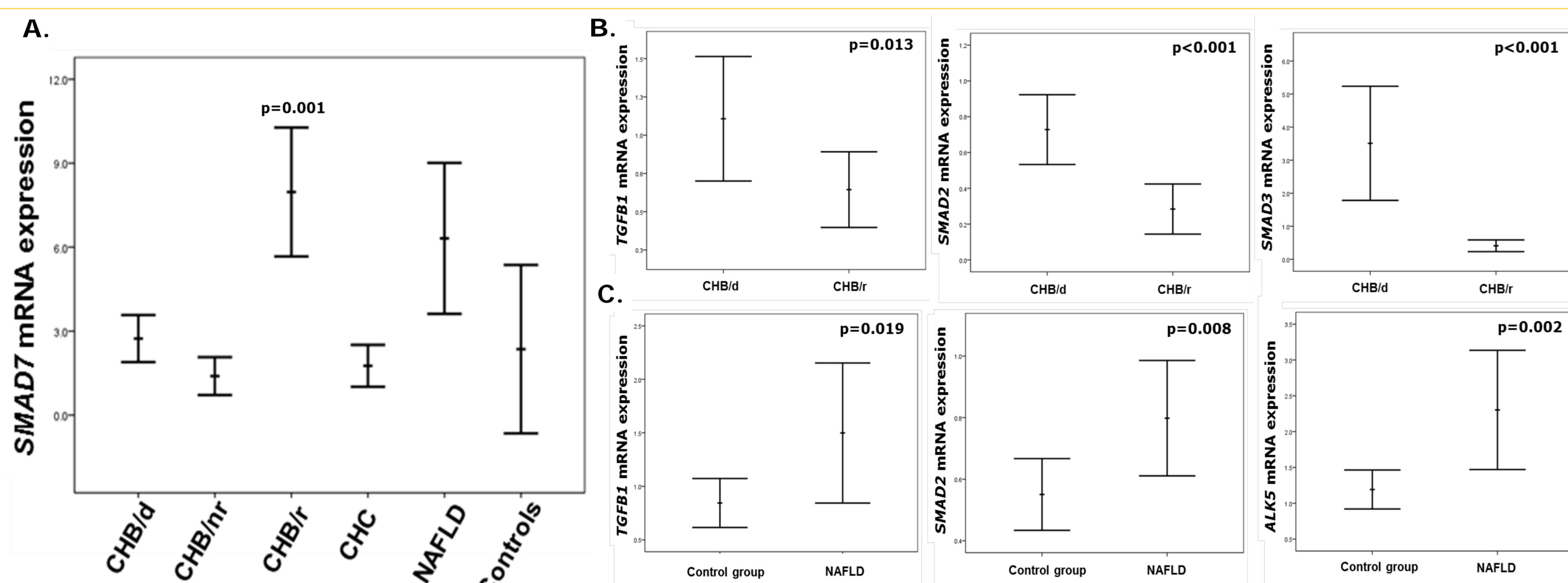


Figure 1. Error bar diagrams presenting the expression of genes, for which a significant alteration of their mRNA levels was observed. p values in each diagram refer to Mann-Whitney U test.

Patients with CHB/r exhibited a significant increase of *SMAD7* mRNA expression (**Fig.1A**) and reduced levels of *TGFb1*, *SMAD2*, *SMAD3*, and *CTGF* ($p=0.010$) as compared to CHB/d patients (**Fig.1B**). This pattern of expression of *SMAD7* was similar with that observed in patients with NAFLD, a disease characterized rarely by a fibrotic process (**Fig.1A**).

SMAD7 expression was also found increased in NAFLD patients as compared to the control group including CHB/d, CHB/nr and CHC patients ($p=0.001$). Moreover, NAFLD patients were presented with elevated mRNA levels of *TGFb1*, *SMAD2*, *ALK4*, and *SMAD4* ($p<0.001$) (**Fig.1C**).

Considering the intensity of inflammation, *SMAD7*, *ALK5*, *INHBC*, and *ALK4* exhibited significant increased expression from absent to minimal inflammation with a gradual reduction as inflammation exacerbates (**Fig.2**).

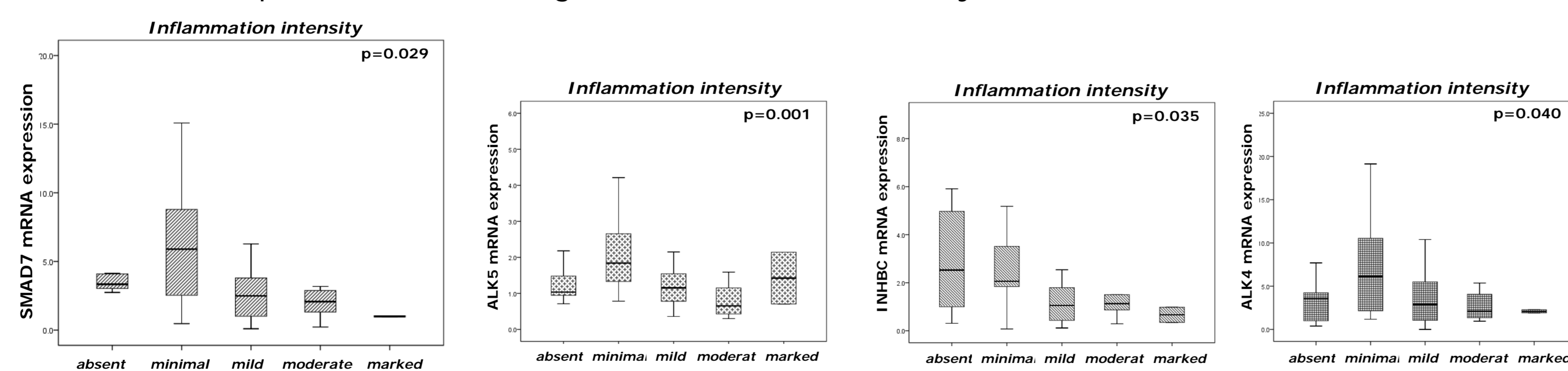


Figure 2. Boxplot diagrams presenting the expression of mediators of the TGFb/Activin signaling pathway according to the intensity of liver inflammation. p values in each diagram refer to Kruskal-Wallis H test.

Conclusion

Our data indicate that in cases with low grade fibrosis, such as NAFLD (characterized by a lower incidence of severe liver complications and fibrosis progression) and CHB/r, *SMAD7* overexpression might be a mechanism limiting the fibrogenic effect of TGFb suggesting that its induction may provide a target for novel therapeutic approaches.

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